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Exploring β -Lactamases genes and Biofilm formation in *E. coli*

isolated from Wastewater, Morocco.

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Introduction

The rise of antibiotic resistance poses a global public health challenge. Wastewater environments, often overlooked, serve as reservoirs for antibiotic-resistant bacteria, fostering the dissemination of resistance genes and posing health risks through various exposure routes. Our study, conducted in Morocco, focuses on the phenotypic and genotypic characterization of antibiotic resistance and biofilm formation in *Escherichia coli* (*E. coli*) strains isolated from wastewater samples.

Methods

Sampling sites: Over a period of two months, from the 1st February to 31st March 2023, wastewater samples were systematically collected from two distinct sites, **Anza** (Latitude: 30.45431; Longitude: -9.65362) and **Oued Lahouar** (Latitude: 30.40016; Longitude: -9.60008), situated in the Agadir city, Morocco. Most of the sampling sites were impacted by pollution from agricultural, industrial, and domestic origins (**Fig. 1**).

Purpose of the study

This study underscores the pivotal role of wastewater in the perpetuation and dissemination of antibiotic resistance, emphasizing the necessity for nuanced wastewater management strategies to curb resistance propagation and safeguard public health.



Fig. 1: Sampling sites

- Antimicrobial Susceptibility Testing was performed by the diffusion method on gelose MH (BioRad), according to the recommendations of CASFM/EUCAST 2022.
- Isolation and identification of bacterial strains: Isolation of *E. coli* strains (n=14) involved the use of a chromogenic medium Brilliance UTI Clarity Agar (Oxoid). Identification was achieved through biochemical and 16S rRNA methods.
- **Biofilm formation on polystyrene microtiter plates:** *E. coli* isolates were incubated on the microplate at various time points ranging from 2 to 24 hours, the absorbance at 620 nm was measured using a microplate reader (800 TS Microplate Reader).
- Detection of β-lactamases genes: PCR assays were conducted to identify genes encoding resistance to β-lactams (*blactx-M*, *blaTEM*, *blaSHV*).

Results

A high prevalence of resistance to various antimicrobial agent was noted (Fig. 2).

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Assessment of biofilm formation on polystyrene microtiter plates showed that the most strains begin biofilm formation between 4h to 18h of incubation, with a moderate formation (20%) after 4h and 22h (Fig. 3).

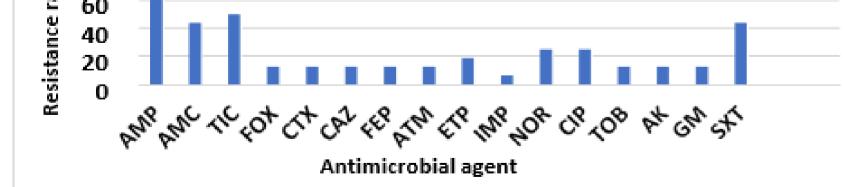


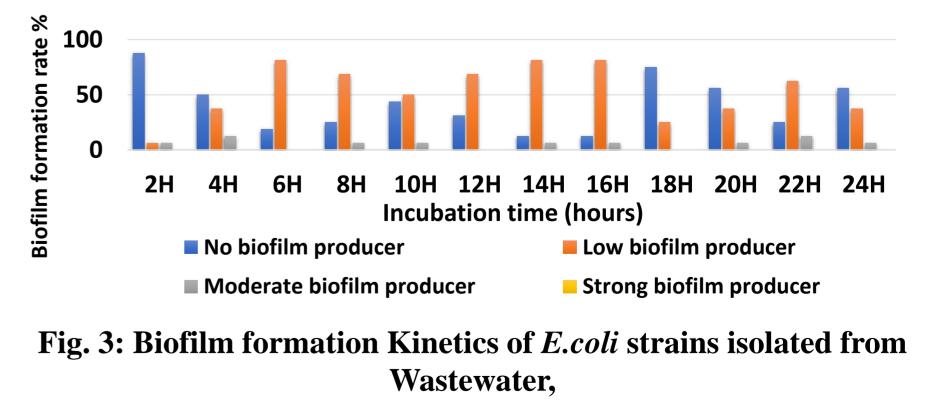
Fig. 2: Antibiotic resistance rate of *E.coli* isolated from Wastewater.

AMP: Ampicillin; AMC: Amoxicillin/clavulanate; TIC: Ticarcillin; FOX: Cefoxitin; CTX: Cefotaxime; CAZ: Ceftazidime; FEP: Cefepime; ETP: Ertapenem, IMP:
Imipenem; NOR: Norfloxacin; CIP: Ciprofloxacin; TOB: Tobramycin; AK: Amikacin; GM: Gentamicin; SXT: co-tromoxazole.

✤ Genetic analysis revealed the presence of *blactx-M*1 in one isolate (Table 1).

Table 1: β-lactamases genes detected in *E.coli* isolated from Wastewater.

Code	Isolate	City	β-lactamases	Antibiotic resistance profile
			genes	
Ec	E.coli	Agadir	blactx-M1	AMP, AMC, TIC, CTX,
A8				CAZ, FEP, ATM



Conclusion

Our findings highlight the critical role of wastewater as a reservoir for antibiotic resistant bacteria emphasizing the need for tailored wastewater management strategies to mitigate resistance dissemination and protect public health.